

バイオインフォマティクスへの招待 ～高速シーケンサーと RNA-Seq～（講義・実習）

Introduction of Bioinformatics - Next Generation Sequencer and RNA-seq

(Lecture/Practice in Japanese)

門田 陽介・芦原 貴司（情報総合センター・医療情報部）

Yosuke Kadota, Takashi Ashihara (Information Technology and Management Center)

この講義・実習では、高速シーケンサーによる塩基配列の読み取りの原理を概説し、得られた膨大な塩基配列データから遺伝子の発現解析（RNA-Seq）を行う手法について、実際にマルチメディアセンターの PC を各自が使ってハンズオンで学ぶ。

We will show you outline the principles of sequence reading by high-speed sequencers, and let you know how to analysis gene expression (RNA-Seq) from the huge amount of sequence data obtained from high-speed sequencers, by using the PCs in the Multimedia Center by yourselves.

# バイオインフォマティクスへの招待

～高速シーケンサーとRNA-Seq～



## What is RNAseq?

- Sequencing of RNA (mainly mRNA) by High-speed sequencer
- Quantitative analysis of gene expression
- Compare with qPCR
  - gene expression can be analyzed comprehensively.
- Splicing analysis is also available

## Examples of analysis

- Comparison of differences in gene expression in normal and cancerous tissues
  - Identification of up- or down-regulated genes and elucidation of disease mechanisms
- Comparison of splicing differences between normal and cancerous tissues
  - Identification of used variants and elucidation of disease mechanisms

## Flow of RNAseq analysis



## Preparation of the environment required for RNAseq

- Installation of software required for analysis
  - Free UNIX-based software
    - Linux or Mac

## WSL(Windows Subsystem for Linux)

- Windows Subsystem for Linux (WSL) is a feature of Windows that allows you to run a Linux environment on your Windows machine, without the need for a separate virtual machine or dual booting. WSL is designed to provide a seamless and productive experience for developers who want to use both Windows and Linux at the same time.



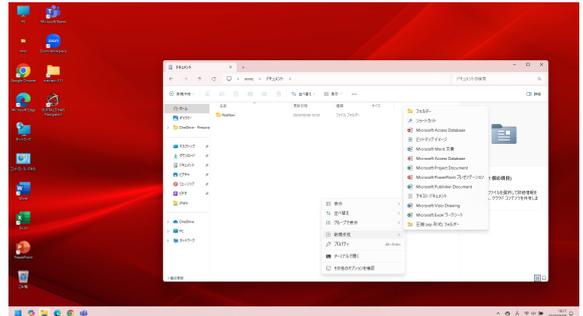
Microsoft Learn  
<https://learn.microsoft.com/en-us/windows/wsl/about>

## CUI (Character User Interface)

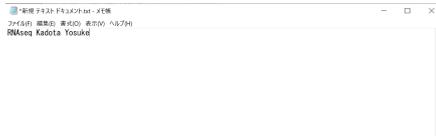


- A method of operating a computer through character input using a keyboard, which allows the user to interact with the computer in a character-based manner.

## GUI (Graphical User Interface)



## GUIとは

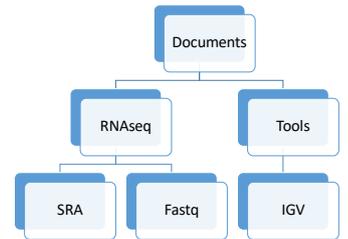


Newtext.txt

## Let's operate by CUI



- Directory (folder) operations
  - cd : Moving Directories
  - pwd : Show current directory
  - ls : Listing of folders and files
  - mkdir : create folder



## Pwd : Show current directory



\$ pwd

```
(base) kadota@Kadota-Lenovo-WS2:~$ pwd
/home/kadota
(base) kadota@Kadota-Lenovo-WS2:~$
```

## cd



\$ cd ..

```
(base) kadota@Kadota-Lenovo-WS2:~$ cd ..
(base) kadota@Kadota-Lenovo-WS2:~/home$ pwd
/home
```

## Ls : Listing of folders and files

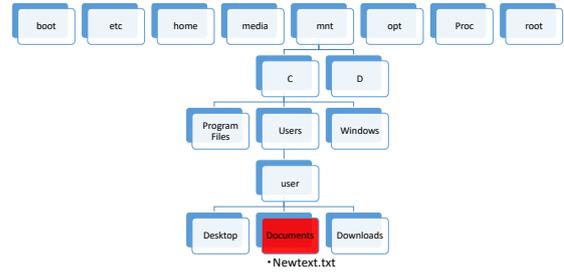


\$ ls

```
(base) kadota@Kadota-Lenovo-WS2:~/home$ cd ..
(base) kadota@Kadota-Lenovo-WS2:/$ pwd
(base) kadota@Kadota-Lenovo-WS2:/$ ls
bin  etc  lib  libx32  mnt  root  snap  tmp
boot home lib32 lost+found opt  run  srv  usr
dev  init lib64 media  proc sbin sys  var
(base) kadota@Kadota-Lenovo-WS2:/$
```

\$ ls -l  
\$ ll

## Let's operate by CUI



## ls



```
(base) kadota@Kadota-Lenovo-WS2:~/Documents$ ls
Newtext.txt
```

## Cat : View all of the text files



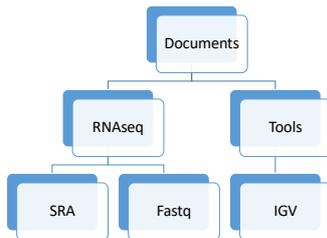
\$ cat Newtext.txt

## mkdir : create folder

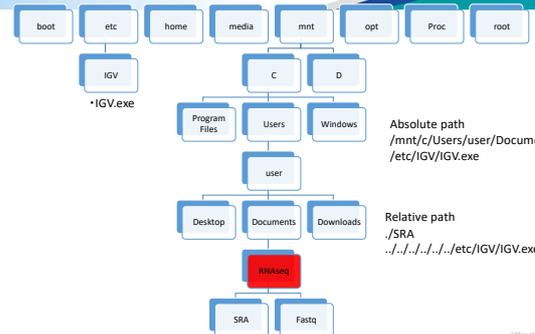


\$ mkdir RNAseq

```
~/Documents$ mkdir RNAseq
~/Documents$ mkdir Tools
~/Documents$ cd RNAseq/
~/Documents/RNAseq$ mkdir SRA Fastq
~/Documents/RNAseq$ cd ..
~/Documents$ cd Tools/
~/Documents/Tools$ mkdir IGV
```



## absolute path and relative path



Absolute path  
/mnt/c/Users/user/Documents/RNAseq/SRA  
/etc/IGV/IGV.exe

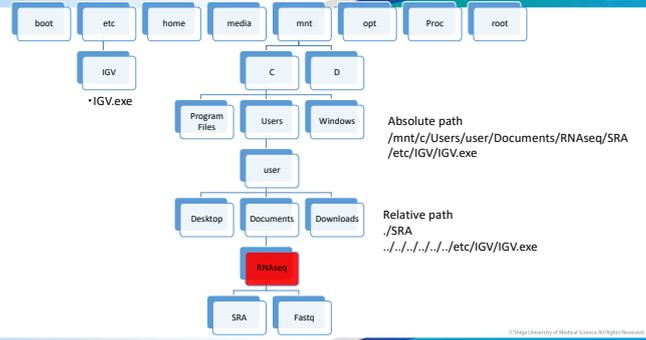
Relative path  
./SRA  
../-./-./-./etc/IGV/IGV.exe

## CUIで操作してみよう

### • ファイルの操作

- mv : Moving files and folders
- cp : Copying files and folders
- cat : View all of the text files
- less : View some of the text files

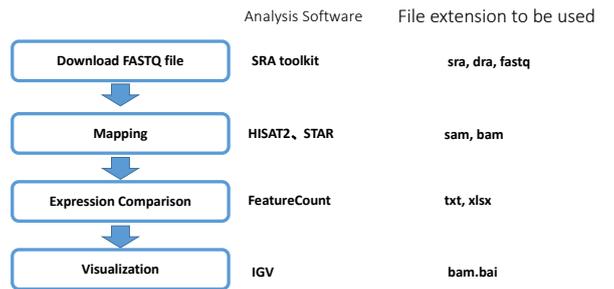
## absolute path and relative path



## less : View some of the text files

\$ less ../Newtext.txt

## RNAseq flowchart



## mapping

GCTATGAAAGTT

CCGGAATTGGAC

ACGGTAACCGTAGCTATGAAAGTT CCGTAAGTACGTTAACCGGAATTGGACCACTAGTC

Genome sequence

## mapping



Genome sequence from ensembl





## FASTQの中身を確認



```
$ less L-1_P1.fastq.gz
```

```
@MG00HS09:723:C9A15ACXX:7:1101:1483:1919 1:N:0:CGATGT
NGACCCGCTGAATTTAAGCATATTAGTCAGCGGAGGAAAAGAACTAACCA
+
#11B?D@8DAF?DGGECHFDF?4ACFB?GEGGB6)?69DE;FGE=4@F)=C
```

- 1 : @Sequence ID and additional information
- 2 : Nucleotide sequence
- 3 : + Sequence ID and additional information
- 4 : sequence quality

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## ASC2

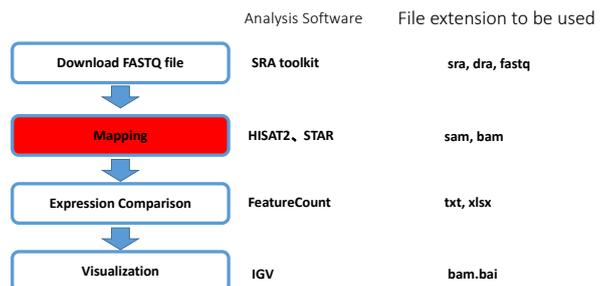


33!	45-	57 9	69E	81Q
34"	46.	58:	70F	82R
35#	47/	59;	71G	83S
36\$	48 0	60<	72H	84T
37%	49 1	61=	73I	85U
38&	50 2	62>	74J	86V
39'	51 3	63?	75K	87W
40(	52 4	64@	76L	88X
41)	53 5	65A	77M	89Y
42*	54 6	66B	78N	90Z
43+	55 7	67C	79O	
44,	56 8	68D	80P	

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ASCII文字	ASCIIコード	クオリティスコア	塩基が間違っている確率 (P)	ASCII文字	ASCIIコード	クオリティスコア	塩基が間違っている確率 (P)
!	33	0	1.0 (100%)	6	54	21	0.00794 (0.794%)
"	34	1	0.794 (79.4%)	7	55	22	0.00631 (0.631%)
#	35	2	0.631 (63.1%)	8	56	23	0.00501 (0.501%)
\$	36	3	0.501 (50.1%)	9	57	24	0.00398 (0.398%)
%	37	4	0.398 (39.8%)	:	58	25	0.00316 (0.316%)
&	38	5	0.316 (31.6%)	;	59	26	0.00251 (0.251%)
'	39	6	0.251 (25.1%)	<	60	27	0.00199 (0.199%)
(	40	7	0.199 (19.9%)	=	61	28	0.00158 (0.158%)
)	41	8	0.158 (15.8%)	>	62	29	0.00126 (0.126%)
*	42	9	0.126 (12.6%)	?	63	30	0.00100 (0.1%)
+	43	10	0.100 (10%)	@	64	31	0.000794 (0.0794%)
,	44	11	0.0794 (7.94%)	A	65	32	0.000631 (0.0631%)
-	45	12	0.0631 (6.31%)	B	66	33	0.000501 (0.0501%)
.	46	13	0.0501 (5.01%)	C	67	34	0.000398 (0.0398%)
/	47	14	0.0398 (3.98%)	D	68	35	0.000316 (0.0316%)
0	48	15	0.0316 (3.16%)	E	69	36	0.000251 (0.0251%)
1	49	16	0.0251 (2.51%)	F	70	37	0.000199 (0.0199%)
2	50	17	0.0199 (1.99%)	G	71	38	0.000158 (0.0158%)
3	51	18	0.0158 (1.58%)	H	72	39	0.000126 (0.0126%)
4	52	19	0.0126 (1.26%)	I	73	40	0.000100 (0.01%)
5	53	20	0.0100 (1.0%)				

## RNAseq flowchart



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## Mapping



- HISAT2
  - `hisat2 -p [number of CPU] -x [genome index] -U [input file] -S [output file]`
  - Download Index file of the genome
    - <http://daehwankimlab.github.io/hisat2/download/#m-musculus>
  - Mapping by using hisat2. Make sam file.
- ```
$ hisat2 -p 8 -x HISATindex/genome -U fastq.gz/L-1_P1.fastq.gz -S sam/L1.sam
```

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## SAMファイル



- QNAME: Query name. Read identifier (usually the read ID).
- FLAG: Bit flag indicating alignment information. It has binary information such as read mapping status and pair information.
- RNAME: Reference name. The name of the reference sequence to which the read is aligned (e.g., chromosome name).
- POS: Leftmost position of 1-base. The starting position of the read mapping. MAPQ: Mapping quality. A score indicating the reliability of the alignment.
- CIGAR: CIGAR string. Indicates the alignment pattern of the reads (e.g., '76M' means that 76 bases were matched).
- RNEXT: The reference name to which the next read is aligned. In the case of paired-end sequencing, the reference sequence to which the other read is aligned.
- PNEXT: The leftmost position of one base of the next read. The starting position of the partner of the paired-end read.
- TLEN: Template Length. Insertion size between paired-end reads.
- SEQ: Read Sequence. Actual nucleotide sequence.
- QUAL: Quality score. Read sequence quality expressed in ASCII characters.

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## Convert sam file to bam file

- samtools

Convert sam file to bam file

```
$ samtools view -@ 8 -bS [input sam file] > [output bam file]
```

Sorting bam files

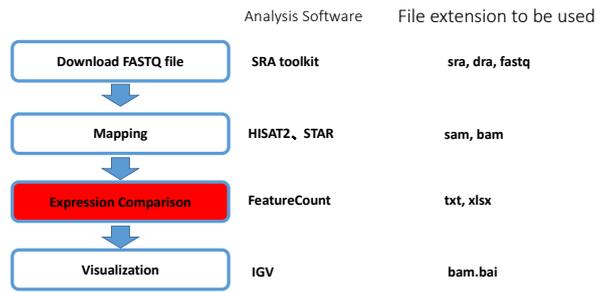
```
$ samtools sort -@ 8 -o [output sorted bam file] [input bam file]
```

Create an index of bam files

```
$ samtools index [input sorted bam file]
```

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## RNAseq flowchart



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## Expression Comparison

- FeatureCounts

```
featureCounts -T [number of CPU] --extraAttributes gene_name -a [GTF file] -o [output file] [input file1] [input file2] [input file3]
```

- Download GTF (gene feature format)

- <https://www.ensembl.org/info/data/ftp/index.html>

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## Expression Comparison

- FeatureCounts

- Count the number of reads mapped to each gene

```
$ featureCounts -T 8 --extraAttributes gene_name -a GTF/Mus_musculus.GRCm38.93.sorted.gtf -o counts_result.txt bam_sort/L1.sort.bam
```

```
$ featureCounts -T 8 --extraAttributes gene_name -a GTF/Mus_musculus.GRCm38.93.sorted.gtf -o counts_result.txt bam_sort/L1.sort.bam bam_sort/L2.sort.bam bam_sort/L3.sort.bam bam_sort/L4.sort.bam bam_sort/L5.sort.bam bam_sort/L6.sort.bam bam_sort/L7.sort.bam bam_sort/L8.sort.bam bam_sort/L9.sort.bam
```

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## Expression Comparison

- Gene expression  $\neq$  Number of reads mapped on the gene
- Longer genes have more reads mapped (intergenic bias)
- The more samples (amount of cDNA), the more reads mapped (sample bias)
- These biases need to be corrected before comparing expression levels.

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## Expression Comparison

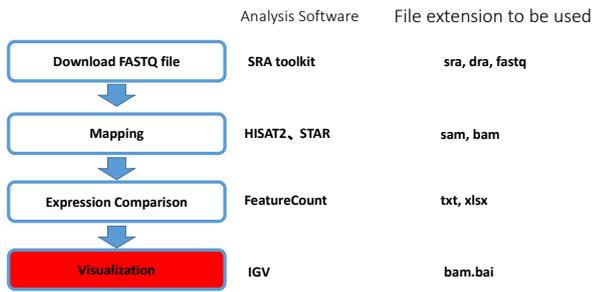
- Excel
  - TPM (transcripts per million)
  - $q_i$  is the number of reads mapped
  - $l_i$  is the transcript length

$$A_i = \frac{q_i}{l_i} * 10^3$$

$$TPM_i = A_i * \frac{1}{\sum_j A_j} * 10^6$$

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## RNAseq flowchart

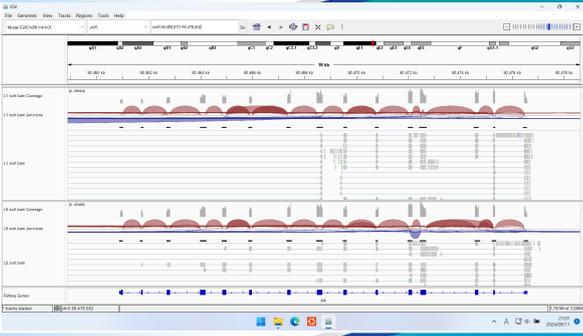


## Visualization

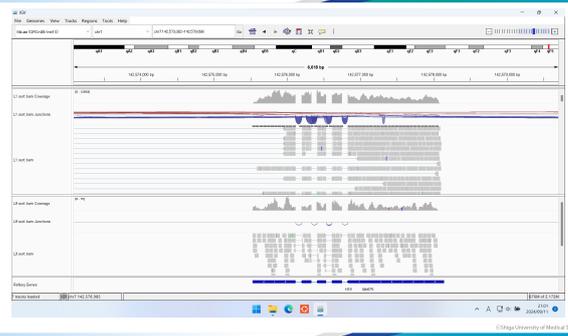


- IGV (Integrative Genomics Viewer)

## Visualization



## Visualization



## Visualization

